

Chapter 5

Nutrition as a Key Factor for Cephalopod Aquaculture

Juan Carlos Navarro, Óscar Monroig and António V. Sykes

Abstract Cephalopods are fast-growing animals, active swimmers and top predators, which require substantial amounts of food. As such, they show high metabolic rates dependent on a carnivorous diet, thus hypothetically linked to a predominant amino acid metabolism. Their body composition is mainly constituted by high levels of total protein, and their lipids, although quantitatively low, reveal the presence of substantial amounts of long-chain polyunsaturated fatty acids. All in all, little is known about their nutritional requirements, especially during the early stages, very prone to high mortalities under culture. This chapter is a brief account of key information concerning relevant points linked to the nutritional requirements that cephalopods have for proteins, lipids, carotenoids, carbohydrates, minerals and vitamins. Moreover, some considerations on populational metabolism are also presented.

Keywords Amino acids · Carbohydrates · Fatty acids · Lipids · Nutrition · *Octopus vulgaris* · Populational metabolism · Proteins · *Sepia officinalis*

5.1 Introduction

Cephalopods potential for aquaculture production was acknowledged by several researchers in the last decades of the twentieth century (e.g. Hanlon 1987; Boucaud-Camou 1989, 1990; Hanlon et al. 1991; Barnabé 1996). Nonetheless, several

J. C. Navarro (✉) · Ó. Monroig
Instituto de Acuicultura Torre de la Sal (IATS-CSIC), 12595 Ribera de Cabanes,
Castellón, Spain
e-mail: jcnavarro@iats.csic.es

Ó. Monroig
Institute of Aquaculture, School of Natural Sciences, University of Stirling,
Stirling FK9 4LA, Scotland, UK
e-mail: oscar.monroig@stir.ac.uk

A. V. Sykes
CCMAR-CIMAR L.A., Centro de Ciências do Mar do Algarve, Universidade do Algarve,
Campus de Gambelas, 8005-139, Faro, Portugal.
e-mail: asykes@ualg.pt

bottlenecks remain and are impeding the transition of technology from pilot to full-scale, namely the existing knowledge on cephalopod nutrition. Nutrition is a key factor for proper growth and survival under captive conditions and mass culture. This chapter picks up from the last review on this subject, made by Lee (1994). Firstly, it presents a nutritional approach, which is mostly based on the biochemical composition of both cephalopods and preys, and finalizes with a metabolic hypothesis, which considers other variables such as enzymes, geographical adaptation and stress. Due to the low amount of existing information for most cephalopod species, this chapter mainly focuses on nutritional studies conducted in *Sepia officinalis* and *Octopus vulgaris*, arguably the cephalopod species of greatest commercial interests.

5.2 Proteins

Proteins are the most abundant macronutrient in cephalopods (Zlatanov et al. 2006) and, as stated by Lee (1994), large protein and amino acid contents in the diet of cephalopods are required for sustaining growth and fulfilling energy demands. According to Lee (1994), cephalopods efficiently absorb, digest and utilize dietary proteins that are further used for locomotion, structural support, energy source, oxygen transport and osmoregulation. Cephalopods display high rates of protein synthesis and retention, and low rates of protein degradation (Houlihan et al. 1990; Moltschanivskyj and Carter 2010). In order to evaluate the dietary requirements of the common octopus *O. vulgaris* during the fast-growing stages, the total and free amino acid (FAA) composition of paralarvae and juveniles was determined by Villanueva et al. (2004). Similar to the amino acid content found in the mantle of juveniles of *O. vulgaris*, *S. officinalis* and *Loligo vulgaris* by Zlatanov et al. (2006), these authors found that glutamate and aspartate were the most abundant nonessential amino acids (NEAAs) in *O. vulgaris* paralarvae, with lysine, leucine and arginine accounting for nearly half of the essential amino acids (EAAs). Interestingly, arginine was the most abundant FAA in paralarvae, possibly indicating its further use for octopine formation produced during an anaerobic work (Baldwin et al. 1976; Hochachka et al. 1976, 1983; Storey and Storey 1979; Storey et al. 1979; Hochachka and Fields 1982) or an active metabolism for energy production and biosynthesis of other amino acids. On the other hand, cephalopods might use proline during oxidative metabolism, either as energy source or as means for augmenting the Krebs cycle (Hochachka and Fields 1982). Both arginine and proline are potentially interconvertible through glutamate and ornithine (Mommensen et al. 1982).

In a general way, there is already information on the amino acids of *O. vulgaris* prey species, such as for *Artemia* spp. (Aragão et al. 2004) and crustaceans such as the spider crab (Andrés et al. 2010), as well as for raw materials used for prepared diets (Valverde et al. 2013). Cephalopod's growth is primarily an increase in body muscle mass by protein synthesis and accretion, and individuals display very high growth rates (especially at the paralarvae and hatchling stages), which means that they have a high dietary requirement for amino acids. In addition, these high growth

rates are most probably due to highly efficient ingestion, digestion (Boucher-Rodoni et al. 1987) and assimilation of protein (Domingues et al. 2005), which have to be supplied by a diet with balanced levels of amino acids, despite the capacity of some cephalopods to perform integumental amino acid uptake from seawater (de Eguileor et al. 2000; Villanueva et al. 2004). Recent results point to the possibility that cuttlefish might not be able to use protein that has been denaturated (Domingues et al. 2009) and to the favour on amino acid use through the pyruvate and tricarboxylic acid pathways in detriment of the ketogenic pathway in starving *O. vulgaris* (García-Garrido et al. 2012). Given its importance for cephalopods metabolism (Lee 1994), there is a need for characterization of the amino acids pool (these should be predominantly used as metabolic fuel, but they should also be utilized for body protein synthesis) at different life stages of different species and preys used for cephalopod rearing. The latter assumes greater importance since marine fish larvae seem to have a lower capacity to digest and absorb complex proteins than juvenile fish (Conceição et al. 2010), and display high amino acid requirements for protein deposition, turnover and catabolism to attain rapid growth (Rønnestad et al. 2003).

In a similar way to demersal fish eggs (Rønnestad et al. 1999), the FAA and protein amino acid pools of *S. officinalis* wild and culture eggs are largely dominated by taurine (more than 50%; Sykes et al. unpublished data). Taurine is an amino acid analogue that is not incorporated into protein, but well known for its multiple roles that include inotropy of high and low calcium, modulator of neuron excitability, resistance to anoxia and hypoxia, bile salt synthesis and simulation of glycolysis and glycogenesis (Huxtable 1992). Taurine is also abundant during planktonic stages of the common octopus where it might play a role in osmoregulation (Villanueva et al. 2004). Nonetheless, its effect on metabolism and growth performance in a fish like gilthead seabream was only related to the increasing methionine availability for several important physiological purposes (Pinto et al. 2013).

5.3 Lipids

Little is known about the lipid requirements of cephalopods, apart from the general information drawn from the analysis of their body composition and assumptions made from their feeding habits. It is then difficult to separate early stages from adult requirements, or establish specific differences. Efforts devoted to the culture of certain species like *O. vulgaris* and *S. officinalis* have provided some information.

Very low levels of total lipids (TL) are present in the mantle of adults (Boucaud-Camou 1990; Sykes et al. 2009b) and hatchlings (Navarro and Villanueva 2000, 2003). This fact, combined with their poor capacity for mitochondrial lipid oxidation (O'Dor et al. 1984; Hochachka 1994), has somehow put aside research on cephalopod lipid nutrition until recently, when research on the causes of the massive mortalities encountered during the culture of early stages of merobenthic species has brought back the protagonism of these essential components on cephalopod nutrition, perhaps with an overemphasis on quantitative rather than qualitative points of view, given the poor essential lipid composition of the live preys commonly used

in aquaculture. Besides, other particular aspects, like the lipid-rich nervous system of hatchlings of *O. vulgaris* paralarvae representing approximately one quarter of the animal's fresh weight (Packard and Albergoni 1970), suggest the importance of lipids for suitable growth during planktonic life. Navarro and Villanueva (2000, 2003) made the first approach towards the study of lipid requirements in early stages of cephalopods to conclude that a nutritional imbalance in the lipid and fatty acid (FA) profile of the artificial feeding protocol may be responsible for the high mortalities encountered. In particular, *O. vulgaris* should require feeding on low-lipid preys, rich in polar lipids (PL), long-chain polyunsaturated fatty acids (PUFA) and possibly cholesterol (Navarro and Villanueva 2000, 2003; Okumura et al. 2005; Seixas et al. 2008). This closely resembles the composition of a 'natural' diet based on crustacean larvae and other marine planktonic organisms like copepods, but is far from the typical composition of the enriched *Artemia* spp. in any of its forms. The picture is even more complicated after evidences pointing at the paralarvae as specialist predators, contrary to the general concept by which they had been often regarded as generalist predators (Roura et al. 2012).

On the other hand, cuttlefish are unable to store lipids in the digestive gland (Fluckiger et al. 2008) and require high levels of phosphatidylcholine, phosphatidylethanolamine (PE) and cholesterol in their diets (Almansa et al. 2006). This conclusion was drawn by the latter authors based on data of whole animal (Navarro and Villanueva 2000), cuttlefish mantle (Sinanoglou and Miniadis-Meimaroglou 1998, 2000) and also of prey lipid content (Domingues et al. 2003; Domingues et al. 2004). It is, however, interesting to verify that these same lipid classes have the highest levels throughout cuttlefish wild and culture egg embryonic development and that, although there is a difference in the amount of TL of eggs from different geographical locations (Sykes et al. 2009a), there is maintenance on the amount of TL until hatching (Bouchaud and Galois 1990). According to Bouchaud and Galois (1990), the egg-yolk lipids correspond to 14% dry weight in eggs and 15% in hatchlings (mainly phospholipids that may be used for energetic purposes).

However, in the case of octopus, up to now, there is no paralarval food from the aquaculture artificial food chain that can compare with the lipid composition of natural live food, and every effort has to be made to try to increase the essential long-chain PUFA and PL content of live preys. In fact, it is not only the bulk provision of essential lipids that is important but also the adequate lipid form (Guinot et al. 2013). This scenario may be more important for early stages, and even paramount in some animal groups like cephalopods, whose FA composition is essentially constituted by palmitic acid (16:0), stearic acid (18:0), docosahexaenoic acid (22:6n-3, DHA) and eicosapentaenoic acid (20:5n-3, EPA), the latter two being essential long-chain highly unsaturated fatty acids (HUFA) for marine organisms (Sinanoglou and Miniadis-Meimaroglou 1998; Navarro and Villanueva 2000; Passi et al. 2002; Ozyurt et al. 2006; Zlatanos et al. 2006). However, the most recent results from Seixas et al. (2010) do not completely exclude the importance of DHA for the successful rearing of *O. vulgaris* paralarvae but seem to point that maybe other n-3 HUFA would be more vital. Very interesting results from Quintana (2009) have shown that EPA is particularly important in paralarvae lipids, with 1:1 EPA:DHA proportions

in the PE, as opposed to the 1:2 proportion generally reported for marine fish. Likewise, EPA is higher in shrimp than fish diets tested as cuttlefish food (Domingues et al. 2003) and its importance for normal growth was proposed by Almansa et al. (2006). The latter authors also indicated that cuttlefish metabolism of both EPA and DHA might be somehow different from fish, where in general the reduction of EPA and DHA sources implies a reduction of the level of these FA in several tissues (Sargent et al. 1995). In addition, Domingues et al. (2003) indicated a 2:1 EPA:DHA proportion in the prey supplied to hatchlings and, in the Ferreira et al. (2010) work, cuttlefish juveniles attaining the best growth and survival rates were fed shrimp with a similar EPA:DHA ratio and PUFA content. On the other hand, Koueta et al. (2002) and Perrin et al. (2004) have suggested the importance of these FAs and PUFA in cuttlefish hatchling survival when facing a stressful situation (either lower water quality or improper prey size, respectively). This suggests the importance of PUFA in cephalopod nutrition and may suggest, for example, that in addition to DHA, EPA may play an important role in the brain and visual system of cephalopods.

A closer look at the available results on the FA profile of *O. vulgaris* paralarvae (Navarro and Villanueva 2000; Miliou et al. 2006) reveals that, apart from the high levels of DHA, 16:0 and EPA, arachidonic acid (20:4n-6, ARA) is one of the most abundant FA, with surprisingly high values of 18:2n-6 and other n-6 FA being found in marine species. From these and other findings in other marine molluscs (Uki et al. 1986; Dunstan et al. 1996; Durazo-Beltran et al. 2003), it has been suggested that 20:4n-6 might not be essential, since the ability for enzymatic bioconversion from adequate precursors to 20:4n-6 could be present in this species. On the other hand, 18:3n-3 is not present in octopus paralarval tissues (Navarro and Villanueva 2000, 2003; Miliou et al. 2006) although this FA is massively included through the *Artemia* spp. feeding. Moreover, 18:3n-3 may compete with 24:5n-3 for the $\Delta 6$ desaturase in the metabolic route leading to DHA production. Therefore, it might be possible that a diet rich in 18:3n-3 could be affecting the proper production of n-3 essential FA (EFA) of paralarvae. An opposite hypothesis is that the C18 desaturation–elongation pathways of 18:3n-3 and 18:2n-6 may be active in *O. vulgaris* so as to produce physiologically essential n-3 and n-6 FA. To explore these and other aspects of lipid metabolism research on dynamic aspects of lipid nutrition and metabolism by characterizing enzymes involved in lipid biosynthesis in these organisms, i.e. desaturases and elongases, is an invaluable tool. Monroig et al. (2012a) have recently isolated a complementary DNA (cDNA) with high homology to fatty acyl desaturases (Fad) in adult octopus. Functional characterization of this enzyme showed that the octopus Fad exhibited $\Delta 5$ -desaturation activity towards saturated and polyunsaturated fatty acyl substrates. Thus, it efficiently converted 16:0 and 18:0 to 16:1n-11 and 18:1n-13, respectively, and desaturated PUFA substrates 20:4n-3 and 20:3n-6 to 20:5n-3 (EPA) and 20:4n-6 (ARA), respectively. Although the $\Delta 5$ Fad enables common octopus to produce EPA and ARA, the low availability of its adequate substrates 20:4n-3 and 20:3n-6, either in the diet or by limited endogenous synthesis from C18 PUFA, might indicate that EPA and ARA are indeed EFA for this species. Interestingly, the octopus $\Delta 5$ Fad can also participate in the biosynthesis of non-methylene-interrupted diene (NMID) FA, PUFA that

are generally uncommon in vertebrates but have been found previously in marine invertebrates, including molluscs, and now also confirmed to be present in specific tissues of common octopus.

Molecular cloning and functional characterization of a cDNA encoding a putative elongase of very long-chain fatty acids (Elovl), a critical enzyme that catalyzes the elongation of FA including PUFA, in common octopus, suggests its phylogenetic relation to Elovl5 and Elovl2, two elongases with demonstrated roles in PUFA biosynthesis in vertebrates (Monroig et al. 2012b). Functional characterization of the octopus Elovl showed the ability to elongate some C18 and C20 PUFAs, while C22 PUFA substrates remained unmodified. Interestingly, the octopus Elovl *elongates* n-6 PUFA substrates more efficiently than their homologous n-3 substrates, suggesting that n-6 PUFA may have particular biological significance in *O. vulgaris*, as mentioned above, and stressing again the essentiality of long-chain n-3 PUFA, and in particular DHA. Besides, the elongase also plays a pivotal role in the biosynthesis of NMID FA.

Similar studies carried out in other cephalopods like *S. officinalis* show a strict parallelism in terms of both qualitative FA composition (see Navarro and Villanueva 2000; Almansa et al. 2006) and biosynthetic capacity (Monroig et al., unpublished data). This, along with tissue FA composition (Monroig et al. 2012a), emphasizes the importance and essentiality of long-chain FA for cephalopods, and validates the idea of using the information to establish the guidelines of the requirements for coastal cephalopods. The recent manuscript by Valverde et al. (2012) on lipid classes from marine species and meals intended for cephalopod feeding has provided additional information on lipids.

5.4 Carotenoids

Carotenoid deposition has been described in cephalopods at the digestive gland (Fox 1966) and in accessory nidamental glands (Decleir and Richard 1972; Van den Branden et al. 1978; Van Den Branden et al. 1980) of *S. officinalis* and proposed to be designated as sepiaxanthine, but there is scarce information about its physiological role. Carotenoid astaxanthin can also be deposited in the skin of *S. officinalis* when fed on grass shrimp (*Palaemonetes varians*; Almansa et al. 2006) and this prey, used for cuttlefish hatchlings rearing, displays up to ten times more carotenoid content (Domingues et al. 2004). *O. vulgaris* paralarvae seem to be able to deposit part of the canthaxanthin present in *Artemia* spp. and to metabolize this carotenoid to astaxanthin (Rodríguez et al., Universidad de La Laguna, unpublished data). Fisher et al. (1956) reported the existence of vitamin A and, in some species, β -carotene in cephalopods. Taking into account the pro-vitamin A and antioxidant activity of carotenoids (Liñán-Cabello et al. 2002), and the importance of this vitamin in photoreception, growth and development, it would be very interesting to carry out more studies to determine the role of carotenoids in early stages of cephalopods as suggested by Villanueva et al. (2009).

5.5 Carbohydrates

Contrary to proteins and lipids, carbohydrate (CH) nutrition in the common octopus has barely been investigated. Considering their limited abundance compared to other macronutrients (normally under 1% of total dry weight; Vlieg 1984; Kreuzer 1984), it is generally accepted that cephalopods do not have a specific requirement for dietary CH (Lee 1994). While protein and amino acids are the primary energy source for cephalopods, it has been reported that cephalopods including *O. vulgaris* are able to rapidly catabolize dietary CH to account for energy demands in explosive activities such as prey capture and fleeing from predators (Morillo-Velarde et al. 2011). Therefore, CH may significantly contribute to fuelling metabolism in *O. vulgaris* under starvation conditions and, consequently, adequate CH inclusion of diets for octopus culture should not be underestimated. The source of CH utilized in diet formulation also needs to be considered. For instance, while glucose was easily digested by *O. vulgaris* (O'Dor et al. 1984), a recent study has revealed that other sugar types such as starch present in freeze-dried pea may exhibit extremely low digestibility (Morillo-Velarde et al. 2012).

On the other hand, the existence of a CH metabolism has been suggested by Sykes et al. (2009a), due to different temperatures of specific geographical locations that may influence the egg nutritional content and metabolism in *S. officinalis*. The sepia egg yolk is composed by a water-soluble glyco-lipoprotein (Ito et al. 1962; Blanchier 1981). This glyco-lipoprotein has 20% lipids (with 65% phospholipid and minor or no cholesterol contents) and 12.6% of CHs (Ito and Fujii 1962). In fact, cuttlefish eggs from Faro (Portugal) have more CHs than lipids (Sykes et al. 2012). Bouchaud (1991) studied the energetic expenditure of *S. officinalis* during embryonic development and found that eggs with more than 0.075 g displayed a similar amount of energy (1,600 J), which led to a theory on the use of yolk for growth and catabolic purposes that is inversely correlated with temperature (e.g. higher temperature implies higher catabolism). In addition, higher temperatures will imply higher oxygen uptake by the embryo (Wolf et al. 1985), which is attained by the increased water volumes of eggs (Sykes et al. 2009a).

5.6 Minerals

The elemental requirements of *O. vulgaris*, as for cephalopods in general, are poorly understood. Nevertheless, it is accepted that octopuses, as carnivorous species, meet the majority of their elemental requirements from the diet, although direct uptake from the seawater has also been shown to occur through an ion balance mechanism regulated by the digestive gland appendages (Wells and Wells 1989). A literature review on the element concentrations in a series of tissues from cephalopods was reported by Napoleão et al. (2005a). While specific (quantitative) requirements for both essential and nonessential elements have not yet been determined, some

studies aiming to determine the elemental composition of *O. vulgaris* have been conducted as a first approach to establish the dietary requirements in this species (Napoleão et al. 2005a; Napoleão et al. 2005b). Thus, Villanueva and Bustamante (2006) reported the elemental composition of the mature ovary, hatchlings, eggs at different developmental stages, wild juvenile individuals and also paralarvae fed a variety of experimental diets. Generally, S, Na, K, P and Mg were determined as the most abundant elements in *O. vulgaris*. Compared to other cephalopods, hatchlings from *O. vulgaris* contained higher levels of Ag, Cu, Mn, Ni and Zn. Compared to subadults and adults of the common octopus (Seixas et al. 2005), the contents of some nonessential elements, namely Ag, Al, Ba, Cd, Hg and Pb, were lower in hatchlings and reared paralarvae, suggesting an accumulation of such elements during development. Similar accumulation of oligoelements seems to occur in cuttlefish (Lacoue-Labarthe et al. 2008a, b, 2009, 2010a, b, Lourenço et al. 2009). Certain elements with potentially pivotal roles in the octopus and cuttlefish physiology have been studied more extensively. Copper (Cu), a key element in the respiratory function of haemocyanin (Ghiretti 1966; D'Aniello et al. 1986), has been postulated to be required for octopus paralarvae, as suggested by the high levels encountered in paralarvae fed on the natural prey *Maja brachydactyla* zoeae in comparison with *Artemia* spp. nauplii (Villanueva and Bustamante 2006). Moreover, octopuses fed on crustacean-based diets contained increased Cu levels compared to fish-fed octopuses, and therefore hypothesized to partly account for a higher cannibalism rate in the latter (García-García and Cerezo-Valverde 2006). The importance of Cu is also high in *S. officinalis* (Declair et al. 1978), especially during maturation of haemocyanin (Declair and Richard 1970; Declair et al. 1971; Wolf et al. 1980; Beuerlein et al. 2004), and this probably extends to other cephalopods species (Taylor and Anstiss 1999). Similar to octopus, the low content of Cu in prepared diets has been pinpointed to provoke mortality by Castro et al. (1993).

Sulphur (S) is also regarded as an essential element for the common octopus and cuttlefish that needs to be provided in the diet at high quantities to sustain formation of muscular proteins (Lee 1994; Villanueva et al. 2004), vestigial shell (Napoleão et al. 2005b) and chitinized structures such as beaks (Hunt and Nixon 1981). Other elements such as strontium (Sr) and cobalt (Co) appear to be incorporated by the common octopus directly from the seawater and by food intake in cuttlefish. It has been demonstrated that Sr is critical for adequate statolith development and consequently for normal swimming and survival of newly hatched octopus, among other cephalopods (Hanlon et al. 1989). In addition to its potential role as an integral component of vitamin B₁₂, Co has also been pointed out as important in the development of adenochrome, a pigment found in the branchial heart, and therefore with a potential role in excretion (Miyazaki et al. 2001).

The cuttlefish *S. officinalis* has a cuttlebone which is made of calcium aragonite (Hewitt 1975) and may suffer malformation resulting from malnutrition (Boletzky 1974). Since food is the primary pathway for the accumulation of trace elements (Bustamante et al. 2004), the inclusion of calcium in a prepared diet is implicit. In fact, the mineral fraction of cuttlefish accounts for 22–32% in eggs (Sykes et al. 2012) and 6% of dry weight at hatching (Villanueva et al. 2004), which can only be attained in such way.

5.7 Vitamins

Villanueva et al. (2009) reported that information on the vitamin composition of cephalopods is mainly limited to the subadult and adult forms in relation to their edible body portions (mantle and arms) or selected organs (Fisher 1956; Sidwell et al. 1978; Motoe et al. 1997; Cho et al. 2001; Pandit and Magar 1972; Passi et al. 2002; Sikorski and Kolodziejaska 1986). These authors analysed the vitamin content of the early stages of cephalopods as an approach to establish their requirements in culture. Antioxidants such as tocopherols are regarded to be very important for the prevention of lipid oxidation, particularly α -tocopherol since it is degraded to protect PUFA against oxidation in fish larvae (Sargent et al. 1997). In fact, it has been reported that the α -tocopherol requirement may depend on the dietary PUFA level (Stéphan et al. 1995; Halver 2002; Brown et al. 2005). Vitamin A and E (α - and γ -tocopherols) profiles of the European cuttlefish *S. officinalis*, European squid *L. vulgaris* and common octopus *O. vulgaris* laboratory hatchlings and wild juveniles were determined. The vitamin A content in early stages of cephalopods was not much different from that observed in other marine molluscs and fish larvae. Besides, relatively high content of vitamin E was observed in the hatchlings and juveniles. These authors postulated that the high levels of vitamin E are probably associated with the high percentage of oxidation-prone PUFA that are particularly high in paralarval and juvenile cephalopods. In general terms, they concluded that the natural and artificial preys (*Artemia* spp.) of early stages of cephalopods either fulfilled their vitamin requirements directly or provided precursors (i.e. carotenoids) that could be transformed in vitamins.

5.8 Populational Metabolism Differences

Captive cephalopods have a different physiology than those from the wild and their tissues are characterized by thicker mantles, a greater proportion of mitochondria-rich tissue, muscle fibres with smaller mitochondrial cores and fewer small muscle fibres (Pecl and Moltschanivskyj 1999). This suggests a reduced rate of new fibre generation, indicating an alteration to the cellular growth mechanisms and not simply a change in the physiological growth rate observed in several laboratory and field studies (Semmens et al. 2004). Growth registered in captive individuals is usually lower than that estimated or verified for those from nature (an example of this is presented in Chap. 11, in the part of cuttlefish culture in earthen ponds).

As stated by Sykes et al. (2006), the particular metabolism of cephalopods needs to be considered if a successful artificial diet is to be designed. The partitioning of the dietary energy intake by cephalopods in the various types of metabolism, excretion and growth was reviewed by O'Dor and Wells (1987). Bearing that in mind, the nutritional content of the diet must be good enough to sustain the existing cephalopod metabolic costs, so the animal is able to allocate an optimal distribution of surplus

energy to somatic growth and, later, to reproduction (Wells and Clarke 1996). Therefore, the animal has to feed on a diet correctly balanced to its metabolic needs at a given temperature (André et al. 2009). The existing information is resumed to two theories that clash in terms of what the cephalopods use as energy substrate. The first one, by Lee (1994) and Boucher-Rodoni and Mangold (1994), considers that under normal feeding conditions, both growth and energy use the protein fraction as fuel. The second theory, by Storey and Storey (1983) and Hochachka (1994), considers that the CH fraction is used as energy source and the protein fraction is exclusively used for growth. Such contrasting theories may originate from the fact that the same species is physiologically adapted to a given geographical location, with different temperature regimes and food nutritional composition. For instance, *S. officinalis* populations from the English Channel and southern Portugal have been reported to be genetically different (Wolfram et al. 2006). In addition, this species displays a physiological plasticity (Oellermann et al. 2012), which is temperature- and food-dependent (reliant on the lifestyle of cephalopods and their low energy reserves).

Cephalopods have appropriate catabolic pathways to breakdown protein to amino acids to obtain energy (Ballantyne et al. 1981). However, most cephalopods' living strategy is to lay down protein reserves into rapid growth to convert them into gametes (O'Dor et al. 1984; Moltschaniwskyj and Carter 2013). Hypothetically, it would be a waste to partially use amino acids for energy, these being reserves only used in case of starving or at reproduction. This would point to the use of other reserves before protein and amino acids at early stages of life. Fast-growing cephalopods, when fed properly, are extremely efficient in converting food to protein, display low protein degradation and exhibit increased efficiency of retaining synthesized protein (Carter et al. 2009). On the other hand, cephalopods are said to have a limited capacity for lipid oxidation (Ballantyne et al. 1981), and its digestion becomes slow and inefficient due to the lack of emulsifiers (biliary salts) in the digestive tract (Vonk 1962). On the contrary, Moltschaniwskyj and Johnston (2006) have shown that *Euprymna tasmanica* has the ability to digest lipids (very high levels of lipase were found in the digestive gland) but these are not stored in the digestive gland, which indicates the species capacity of using lipids as a source of fuel (Swift et al. 2005). This species is known to have a very sedentary lifestyle (it does not move a lot, so there are not many mantle burst activities) in Southern Australia temperate waters. A similar ability to use lipids had already been displayed by individuals of *S. officinalis* from the English Channel populations, which metabolized lipids instead of protein or CHs when facing prolonged starvation (Castro et al. 1992). On the other hand, Lamarre et al. (2012) observed the mixed use of lipids and protein, and after 8 days the exclusive use of protein in short starvation, in cuttlefish from the Mediterranean Sea. This use of lipids by cuttlefish might be eventually identical, at the cellular level, to what is described by Finn and Dice (2006) in vertebrates.

The higher growth rates observed in cephalopods that live or are cultured in subtropical regions (with high temperatures than those observed in temperate waters) would mean that from a given temperature threshold, cephalopods would use CHs as energy, while at lower temperatures, they would preferably use other sources such as lipids and protein (depending on food availability). The enzyme content of the

different life stages helps to understand the eventual existence of different metabolisms. While the enzyme content of the English Channel population of *S. officinalis* has been reported, for several years and by numerous researchers, those of Faro or Mediterranean populations have never, to the best of our knowledge, been reported. Regarding the first, Boucaud-Camou (1969, 1947) characterized amylase and protease activities in different digestive organs of juvenile cuttlefish and Yim (1978) detected amylase activity in mature cuttlefish. According to Koueta et al. (2000), this activity increases with age, not being present at hatching, which is concomitant with the maturation of the sepia digestive system (Boucaud-Camou et al. 1985). Nonetheless, the use of silage as enrichment for shrimp given as prey to cuttlefish hatchlings promoted an increase of total CH and peptides in the diet and supported 100% survival plus increased growth, when compared with natural diets, and despite the lower content in total protein (Le Bihan et al. 2006). However, these same authors reported an inhibition of amylase activity but higher proteolytic activity.

Higher temperatures promote oxygen-efficient adenosine triphosphate (ATP) production due to limitations in available oxygen (Hochachka 1994; Pörtner 2010). However, most studies on cephalopod metabolism have been performed in fasting animals, where the stressful situation of meeting the energy requirements will promote the use of protein reserves (McCue 2010). In addition, one has to consider that all animals exhibit adaptive biochemical and physiological responses to the lack of food (Wang et al. 2006). This is particularly true regarding cephalopods, most of which inhabit environments in which food availability fluctuates or encounters with appropriate food items might be rare and unpredictable at given geographical locations or seasons.

Houlihan et al. (1990) studied protein metabolism in *O. vulgaris* and concluded that the high growth rates displayed by this species rely on high rates of protein synthesis and high efficiencies of retention of synthesized protein and little protein degradation. According to Oellermann et al. (2012), the European cuttlefish has the capability of adjusting its cellular and mitochondrial energetics over short- and long-term changes of temperature and environmental conditions, which is an evolutionary adaptation of given populations, such as *S. officinalis*. In addition, temperature has a significant effect on oxygen consumption (Grigoriou and Richardson 2009) and energy metabolism of cuttlefish (Mark et al. 2008). Furthermore, temperate cuttlefish (English Channel populations) display a predominant oxidation of proline in systemic heart, while subtropical cuttlefish (Mediterranean populations) exhibit enhanced pyruvate oxidation. The latter is supported by the findings of Ballantyne et al. (1981) on octopine dehydrogenase linking amino acid (arginine) and CH (pyruvate) metabolism, which are said to occur during hypoxic conditions, burst activity or both.

In this sense, cold-adapted metabolism in cuttlefish will show a suppressed CH metabolism, favoring the use of lipids (e.g. results of Koueta et al. (2002); Perrin et al. (2004) and Koueta et al. (2006)) and proline, which are less oxygen efficient (Hochachka 1994). At lower temperatures, amino acids such as glutamate, ornithine or arginine may sustain the supply of succinate (Ballantyne et al. 1981; Hochachka et al. 1983). Cephalopods should have developed an aerobic CH–amino acid metabo-

lism that maximizes ATP yield per unit of consumed oxygen due to the lack of an intracellular myoglobin analogue and the exclusive localization of mitochondria in interfibrillar zones (Hochachka 1994; Ballantyne 2004). CH are catabolized rapidly following ingestion, and are not used during fasting, but mobilized for locomotor activity (O'Dor et al. 1984). According to Hochachka and Fields (1982) and Storey and Storey (1978), glucose metabolism may be coupled to that of proline through glucose conversion to pyruvate, which is oxidized in the Krebs cycle, and proline simultaneously augments the cycle intermediates. These first authors still add that glutamate, proline and most probably arginine can be used directly as gluconeogenic precursors.

5.9 Conclusions

Proteins are the most abundant macronutrient in cephalopods, and large protein and amino acid contents in the diet are required for sustaining growth and eventually fulfilling energy demands. Although low in quantitative terms, lipids are essential in cephalopod nutrition, with long-chain PUFA playing a pivotal role, since the enzymatic machinery unveiled through molecular research points at their inability in the synthesis of these important nutrients. Therefore, the effect of dietary protein/lipid ratios on amino acid absorption efficiency and metabolism needs to be researched.

Cephalopods also require an adequate provision of antioxidants like pro-vitamin A and carotenoids, since the importance of vitamins in photoreception, growth and development must not be neglected. It is generally accepted that cephalopods do not have a specific requirements for dietary CH, but it has been reported that they are able to rapidly catabolize dietary CH to account for energy demands in explosive activities such as prey capture and fleeing from predators, and the existence of a CH metabolism has been established. However, the CH theory presented in this chapter needs to be proven. Finally, it is accepted that octopuses and cuttlefish, as carnivorous species, meet the majority of their elemental requirements from the diet, although direct uptake from the seawater has also been shown to occur through an ion balance mechanism regulated by the digestive gland appendages.

Methods for the determination of possible metabolic pathways have been developed for fish larvae (Conceição et al. 2003; Morais et al. 2004) and these may be adapted for the study of metabolism in cephalopods. Additionally, the use of systems biology modelling to cephalopod nutrition, similar to that published by Hormiga et al. (2010), might help on a faster progress towards our understanding of cephalopods' physiology and nutrition.

References

- Almansa E, Domingues P, Sykes A et al (2006) The effects of feeding with shrimp or fish fry on growth and mantle lipid composition of juvenile and adult cuttlefish (*Sepia officinalis*). *Aquaculture* 256:403–413

- André J, Grist EPM, Semmens JM et al (2009) Effects of temperature on energetics and the growth pattern of benthic octopuses. *Mar Ecol Prog Ser* 374:167–179
- Andrés M, Estévez A, Simeó CG et al (2010) Annual variation in the biochemical composition of newly hatched larvae of *Maja brachydactyla* in captivity. *Aquaculture* 310:99–105
- Aragão C, Conceição LEC, Dinis MT et al (2004) Amino acid pools of rotifers and *Artemia* under different conditions: Nutritional implications for fish larvae. *Aquaculture* 234:429–445
- Baldwin J, Fields JHA, Hochachka PW (1976) Role of octopine dehydrogenase in energy-metabolism of cephalopods. *Proc Australian Biochem Soc* 9:5–5
- Ballantyne JS (2004) Mitochondria: aerobic and anaerobic design—lessons from molluscs and fishes. *Comp Biochem Physiol B* 139:461–467
- Ballantyne JS, Hochachka PW, Mommsen TP (1981) Studies on the metabolism of the migratory squid, *Loligo opalescens*—enzymes of tissues and heart-mitochondria. *Mar Biol Lett* 2:75–85
- Barnabé G (1996) Bases Biologiques et Écologiques de l'Aquaculture. Editorial Acribia SA, Zaragoza
- Beuerlein K, Ruth P, Scholz FR et al (2004) Blood cells and the biosynthesis of hemocyanin in *Sepia* embryos. *Micron* 35:115–116
- Blanchier B (1981) Etude des lipides totaux et des stéroïdes dans la glande digestive et la gonade chez la seiche *Sepia officinalis* L. (Mollusque, Cephalopode). Université de Caen, France
- Boletzky SV (1974) Effects of continuous malnutrition on development of cuttlebone in *Sepia officinalis* L. (Mollusca, Cephalopoda). *Bull Soc Zool France—Evol Zool* 99:667–673
- Bouchaud O (1991) Energy consumption of the cuttlefish *Sepia officinalis* L. (Mollusca: Cephalopoda) during embryonic development, preliminary results. *Bull Mar Sci* 49:333–340
- Bouchaud O, Galois R (1990) Utilization of egg-yolk lipids during the embryonic development of *Sepia officinalis* L. in relation to temperature of the water. *Comp Biochem Physiol* 97B:611–615
- Boucaud-Camou E (1969) Localization of amylase and protease activities in digestive system of *Sepia officinalis* L. *Cr Acad Sci D Nat* 269:2564–2566
- Boucaud-Camou E (1974) Localisation d'activités enzymatiques impliquées dans la digestion chez *Sepia officinalis* L. *Arch Zool Exp Gén* 115:5–27
- Boucaud-Camou E (1989) L'aquaculture des céphalopodes: évaluation et perspectives. *Haliotis* 19:201–214
- Boucaud-Camou E (1990) La seiche, un animal d'avenir. *Peche Maritime* 69:321–329
- Boucaud-Camou E, Yim M, Tresgot A (1985) Feeding and digestion of young *Sepia officinalis* L. (Mollusca: Cephalopoda) during post-hatching development. *Vie Milieu* 35:263–266
- Boucher-Rodoni R, Mangold K (1994) Ammonia production in cephalopods, physiological and evolutionary aspects. *Mar Freshw Behav Physiol* 25:53–60
- Boucher-Rodoni R, Boucaud-Camou E, Mangold K (1987) Feeding and digestion. In: Boyle PR (ed) *Cephalopod Life Cycles*. Academic Press London, p 85–108
- Brown MR, Battaglione SC, Morehead DT, Brock M (2005) Ontogenetic changes in amino acid and vitamins during early larval stages of striped trumpeter (*Latris lineata*). *Aquaculture* 248:263–274
- Bustamante P, Teyssie JL, Danis B et al (2004) Uptake, transfer and distribution of silver and cobalt in tissues of the common cuttlefish *Sepia officinalis* at different stages of its life cycle. *Mar Ecol—Progr Ser* 269:185–195
- Carter CG, Lynch KA, Moltschanivskyj NA (2009) Protein synthesis in a solitary benthic cephalopod, the Southern dumpling squid (*Euprymna tasmanica*). *Comp Biochem Physiol A: Mol Int Physiol* 153:185–190
- Castro BG, Garrido JL, Sotelo CG (1992) Changes in composition of digestive gland and mantle muscle of the cuttlefish *Sepia officinalis* during starvation. *Mar Biol* 114:11–20
- Castro BG, Dimarco FP, Derusha RH et al (1993) The effects of surimi and pelleted diets on the laboratory survival, growth, and feeding rate of the cuttlefish *Sepia officinalis* L. *J Exp Mar Biol Ecol* 170:241–252
- Cho SY, Joo DS, Choi HG, Nara E, Miyashita K (2001) Oxidative stability of lipids from squid tissues. *Fish Sci* 67:738–743
- Conceição LEC, Grasdalen H, Ronnestad I (2003) Amino acid requirements of fish larvae and post-larvae: new tools and recent findings. *Aquaculture* 227:221–232

- Conceição L, Aragão C, Ronnestad I (2010) Protein metabolism and amino acid requirements in fish larvae. In: Cruz-Suarez LE, Ricque-Marie D, Tapia-Salazar M, Nieto-López MG, Villarreal-Cavazos DA, Gamboa-Delgado J (eds) *Avances en Nutrición Acuicola X—Memorias del Décimo Simposio Internacional de Nutrición Acuicola*. Monterrey, México, p 250–263
- D’Aniello A, Strazzullo L, D’Onofrio G, Pischetola M (1986) Electrolytes and nitrogen compounds of body fluids and tissues of *Octopus vulgaris* Lam. *J Comp Physiol* 156:503–509
- De Eguileor ML, Grimaldi A et al (2000) Integumental amino acid uptake in a carnivorous predator mollusc (*Sepia officinalis*, Cephalopoda). *Tissue Cell* 32:389–398
- Decleir W, Richard A (1970) A study of the blood proteins in *Sepia officinalis* L. with special reference to embryonic hemocyanin. *Comp Biochem Physiol* 34:203–211
- Decleir W, Richard A (1972) A study of the orange-red pigment from the accessory nidamental glands of the cephalopod *Sepia officinalis* L. *Biol Jb Dodonaea* 40:188–197
- Decleir W, Lemaire J, Richard A (1971) The differentiation of blood proteins during ontogeny in *Sepia officinalis* L. *Comp Biochem Physiol* 40B:923–930
- Decleir W, Vlaeminck A, Geladi P et al (1978) Determination of protein-bound copper and zinc in some organs of the cuttlefish *Sepia officinalis* L. *Comp Biochem Physiol* 60B:347–350
- Domingues P, Poirier R, Dickel L et al (2003) Effects of culture density and live prey on growth and survival of juvenile cuttlefish, *Sepia officinalis*. *Aquac Int* 11:225–242
- Domingues P, Sykes A, Sommerfield A et al (2004) Growth and survival of cuttlefish (*Sepia officinalis*) of different ages fed crustaceans and fish. Effects of frozen and live prey. *Aquaculture* 229:239–254
- Domingues PM, Dimarco FP, Andrade JP et al (2005) Effect of artificial diets on growth, survival and condition of adult cuttlefish, *Sepia officinalis* Linnaeus, 1758. *Aquac Int* 13:423–440
- Domingues P, Marquez L, López N et al (2009) Effects of food thermal treatment on growth, absorption, and assimilation efficiency of juvenile cuttlefish, *Sepia officinalis*. *Aquac Int* 17:283–299
- Dunstan GA, Baillie HJ, Barrett SM, Volkman JK (2006) Effect of diet on the lipid composition of wild and cultured abalone. *Aquaculture* 140:115–127
- Durazo-Beltrán E, D’Abramo LR, Toro-Vazquez JF et al (2003) Effect of triacylglycerols in formulated diets on growth and fatty acid composition in tissue of green abalone (*Haliotis fulgens*). *Aquaculture* 224:257–270
- Ferreira A, Marquez L, Almansa E et al (2010) The use of alternative diets to culture juvenile cuttlefish, *Sepia officinalis*: effects on growth and lipid composition. *Aquac Nut* 16:262–275
- Finn PF, Dice JF (2006) Proteolytic and lipolytic responses to starvation. *Nutrition* 22:830–844
- Fisher LR, Kon SK, Thompson SY (1956) Vitamin A and carotenoids in certain invertebrates. V Mollusca: Cephalopoda. *J Mar Biol Assoc UK* 35:63–80
- Fluckiger M, Jackson GD, Nichols P et al (2008) An experimental study of the effect of diet on the fatty acid profiles of the European cuttlefish (*Sepia officinalis*). *Mar Biol* 154:363–372
- Fox DL (1966) Pigmentation of Molluscs. In: Yonge CM (ed) *Physiology of Mollusca*. Academic Press Inc., New York, p 240–273
- García-García B, Cerezo-Valverde J (2006) Optimal proportions of crabs and fish in diet for common octopus (*Octopus vulgaris*) on-growing. *Aquaculture* 253:502–511
- García-Garrido S, Hachero-Cruzado I, Rosas C et al (2012) Protein and amino acid composition from the mantle of juvenile *Octopus vulgaris* exposed to prolonged starvation. *Aquac Res* 44:1741–1751
- Grigoriou P, Richardson CA (2009) Effect of body mass, temperature and food deprivation on oxygen consumption rate of common cuttlefish *Sepia officinalis*. *Mar Biol* 156:2473–2481
- Guinot D, Monroig Ó, Navarro JC et al (2013). Enrichment of *Artemia* metanauplii in phospholipids and essential fatty acids as a diet for common octopus (*Octopus vulgaris*) paralarvae. *Aquac Nut* 19:837–844
- Ghiretti F (1966) Molluscan hemocyanins. In: Wilbur KM, Yonge CM (eds) *Physiology of Mollusca*, vol II. Academic Press, London
- Halver JE (2002) The vitamins In: Halver JE, Hardy RW (eds) *Fish Nutrition*, 3rd edn. Academic Press, San Diego, p 61–141

- Hanlon RT (1987) Mariculture. In: Cephalopod Life Cycles. Academic Press Inc., London
- Hanlon RT, Bidwell JP, Tait R (1989) Strontium Is required for statolith development and thus normal swimming behavior of hatching cephalopods. *J Exp Biol* 141:187–195
- Hanlon RT, Turk PE, Lee PG (1991) Squid and cuttlefish mariculture: an updated perspective. *J Ceph Biol* 2:31–40
- Hewitt RA (1975) Analysis of aragonite from cuttlebone of *Sepia officinalis* L. *Mar Geol* 18:M1–M5
- Hochachka PW (1994) Oxygen efficient design of cephalopod muscle metabolism. *Mar Fresh Behav Physiol* 25:61–67
- Hochachka PW, Fields JHA (1982) Arginine, glutamate, and proline as substrates for oxidation and for glycogenesis in cephalopod tissues. *Pac Sci* 36:325–335
- Hochachka PW, Hartline PH, Fields JHA (1976) Octopine as an end product of anaerobic glycolysis in the chambered nautilus. *Science* 195:72–74
- Hochachka PW, Mommsen TP, Storey J et al (1983) The relationship between arginine and proline metabolism in cephalopods. *Mar Biol Lett* 4:1–21
- Hormiga JA, Almansa E, Sykes AV et al (2010) Model based optimization of feeding regimens in aquaculture: Application to the improvement of *Octopus vulgaris* viability in captivity. *J Biotechnol* 149:209–214
- Houlihan DF, Mcmillan DN, Agnisola C et al (1990) Protein synthesis and growth in *Octopus vulgaris*. *Mar Biol* 106:251–259
- Hunt S, Nixon M (1981) A comparative study of protein composition in the chitin-protein complexes of the beak, pen, sucker disk, radula and esophageal cuticle of cephalopods. *Comp Biochem Physiol B* 68:535–546
- Huxtable RJ (1992) Physiological actions of taurine. *Physiol Rev* 72:101–163
- Ito Y, Fujii T (1962) Chemical composition of the egg-yolk lipoproteins. *J Biochem* 52:221–222
- Ito Y, Fujii T, Otake M (1962) On peptide constituent of the egg-yolk lipoproteins. *J Biochem* 52:223–225
- Koueta N, Le CA, Noel B et al (2000) Changes of digestive enzymes during growth of cultured juvenile cuttlefish *Sepia officinalis* L. (Mollusca Cephalopoda). Effect of enriched diet and ration. In: ICES (ed) ICES Annual Science Conference. Brugge, Belgium, p 17
- Koueta N, Boucaud-Camou E, Noel B (2002) Effect of enriched natural diet on survival and growth of juvenile cuttlefish *Sepia officinalis* L. *Aquaculture* 203:293–310
- Koueta N, Alorend E, Noel B et al (2006) Earlier acceptance of frozen prey by juvenile cuttlefish *Sepia officinalis* in experimental rearing: effect of previous enriched natural diet. *Vie Milieu* 56:147–152
- Kreuzer R (1984) Cephalopods: handling, processing and products. *FAO Fish Tech Pap* 254:108
- Lacoue-Labarthe T, Warnau A, Oberhansli F et al (2008a) Differential bioaccumulation behaviour of Ag and Cd during the early development of the cuttlefish *Sepia officinalis*. *Aquat Toxicol* 86:437–446
- Lacoue-Labarthe T, Warnau M, Oberhansli F et al (2008b) First experiments on the maternal transfer of metals in the cuttlefish *Sepia officinalis*. *Mar Pollut Bull* 57:826–831
- Lacoue-Labarthe T, Warnau M, Metian M et al (2009) Biokinetics of Hg and Pb accumulation in the encapsulated egg of the common cuttlefish *Sepia officinalis*: Radiotracer experiments. *Sci Total Environ* 407:6188–6195
- Lacoue-Labarthe T, Le Bihan E, Borg D et al (2010a) Acid phosphatase and cathepsin activity in cuttlefish (*Sepia officinalis*) eggs: the effects of Ag, Cd, and Cu exposure. *ICES J Mar Sci* 67:1517–1523
- Lacoue-Labarthe T, Warnau M, Oberhansli F et al (2010b) Contrasting accumulation biokinetics and distribution of Am-241, Co, Cs, Mn and Zn during the whole development time of the eggs of the common cuttlefish, *Sepia officinalis*. *J Exp Mar Biol Ecol* 382:131–138
- Lamarre SG, Ditlecadet D, McKenzie DJ et al (2012) Mechanisms of protein degradation in mantle muscle and proposed gill remodeling in starved *Sepia officinalis*. *Am J Physiol-Reg I* 303:R427–R437
- Le Bihan E, Perrin A, Koueta N (2006) Influence of diet peptide content on survival, growth and digestive enzymes activities of juvenile cuttlefish *Sepia officinalis*. *Vie Milieu* 56:139–145

- Lee PG (1994) Nutrition of cephalopods: fueling the system. *Mar Fresh Behav Physiol* 25:35–51
- Liñán-Cabello MA, Paniagua-Michel J, Hopkins PM (2002) Bioactive roles of carotenoids and retinoids in crustaceans. *Aquac Nut* 8:299–309
- Lourenço HM, Anacleto P, Afonso C et al (2009) Elemental composition of cephalopods from Portuguese continental waters. *Food Chem* 113:1146–1153
- Mark F, Melzner F, Bock C et al (2008) Thermal effects on cephalopod energy metabolism—a case study for *Sepia officinalis*. *Comp Biochem Physiol A* 150:S171–S171
- McCue MD (2010) Starvation physiology: reviewing the different strategies animals use to survive a common challenge. *Comp Biochem Physiol A* 156:1–18
- Miliou H, Fintikaki M, Tzitzinakis M et al (2006) Fatty acid composition of the common octopus, *Octopus vulgaris*, in relation to rearing temperature and body weight. *Aquaculture* 256:311–322
- Miyazaki T, Nakahara M, Ishii T et al (2001) Accumulation of cobalt in newly hatched octopus, *Octopus vulgaris*. *Fish Sci* 67:170–172
- Moltschaniwskyj N, Johnston D (2006) Evidence that lipid can be digested by the dumpling squid *Euprymna tasmanica*, but is not stored in the digestive gland. *Mar Biol* 149:565–572
- Moltschaniwskyj NA, Carter CG (2010) Protein synthesis, degradation, and retention: mechanisms of indeterminate growth in cephalopods. *Physiol Biochem Zool*: PBZ 83:997–1008
- Moltschaniwskyj MA, Carter CG (2013) The adaptive response of protein turnover to the energetic demands of reproduction in a cephalopod. *Physiol Biochem Zool* 86:119–126
- Mommsen TP, French CJ, Emmett B et al (1982) The fate of arginine and proline carbon in squid tissues. *Pac Sci* 3:343–348
- Monroig Ó,G, Hontoria F et al (2012a) Biosynthesis of essential fatty acids in *Octopus vulgaris* (Cuvier, 1797): Molecular cloning, functional characterization and tissue distribution of a fatty acyl elongase. *Aquaculture* 360–361:45–53
- Monroig Ó,N, Dick JR et al (2012b) Identification of a $\Delta 5$ -like fatty acyl desaturase from the cephalopod *Octopus vulgaris* (Cuvier 1797) involved in the biosynthesis of essential fatty acids. *Mar Biotech* 14:411–422
- Morais S, Conceição L, Dinis MT, Rønnestad I (2004) A method for radiolabeling *Artemia* with applications in studies of food intake, digestibility, protein and amino acid metabolism in larval fish. *Aquaculture* 231:469–487
- Morillo-Velarde PS, Cerezo Valverde J, Serra Llinares RM et al (2011) Energetic contribution of carbohydrates during starvation in common octopus (*Octopus vulgaris*). *J Mollusc Stud* 77:318–320
- Morillo-Velarde PS, Cerezo Valverde J, Hernández MD et al (2012) Growth and digestibility of formulated diets based on dry and freeze-dried ingredients in the common octopus (*Octopus vulgaris*). *Aquaculture* 368–369:139–144
- Motoe K, Ooizumi T, Hayashi S, Kawasaki K (1997) Seasonal changes in the contents of proximate composition, minerals and vitamins of firefly squid. *J Jpn Soc Food Sci Tech* 44:133–139
- Napoleão P, Pinheiro T, Reis CS (2005a) Elemental characterization of tissues of *Octopus vulgaris* along the Portuguese coast. *Sci Total Environm* 345:41–49
- Napoleão P, Reis CS, Alves LC et al (2005b) Morphologic characterization and elemental distribution of *Octopus vulgaris* Cuvier, 1797 vestigial shell. *Nuclear Instrum Meth Physic Research B* 231:345–349
- Navarro JC, Villanueva R (2000) Lipid and fatty acid composition of early stages of cephalopods: an approach to their lipid requirements. *Aquaculture* 183:161–177
- Navarro JC, Villanueva R (2003) The fatty acid composition of *Octopus vulgaris* paralarvae reared with live and inert food: deviation from their natural fatty acid profile. *Aquaculture* 219:613–631
- O’Dor RK, Mangold K, Boucher-Rodoni R et al (1984) Nutrient absorption, storage and remobilization in *Octopus vulgaris*. *Marine Behavioral Physiology* 11:239–258
- O’Dor RK, Wells MJ (1987) Energy and nutrient flow. In: Boyle PR (ed) *Cephalopod life cycles*. Academic Press, London, p 109–133
- Oellermann M, Portner HO, Mark FC (2012) Mitochondrial dynamics underlying thermal plasticity of cuttlefish (*Sepia officinalis*) hearts. *J Exp Biol* 215:2992–3000

- Okumura S, Kurihara A, Iwamoto A et al (2005) Improved survival and growth in *Octopus vulgaris* paralarvae by feeding large type *Artemia* and Pacific sandeel, *Ammodytes personatus*: Improved survival and growth of common octopus paralarvae. *Aquaculture* 244:147–157
- Ozyurt G, Duysak O, Akamca E et al (2006) Seasonal changes of fatty acids of cuttlefish *Sepia officinalis* L. (Mollusca: Cephalopoda) in the north eastern Mediterranean sea. *Food Chem* 95:382–385
- Packard A, Albergoni V (1970) Relative growth, nucleic acid content and cell numbers of brain in *Octopus vulgaris* (Lamarck). *J Exp Biol* 52:539–552
- Pandit AR, Magar NG (1972) Chemical composition of *Sepia orientalis* and *Loligo vulgaris*. *Fish Technol* 9:122–125
- Passi S, Cataudella S, Di Marco P et al (2002) Fatty acid composition and antioxidant levels in muscle tissue of different Mediterranean marine species of fish and shellfish. *J Agric Food Chem* 50:7314–7322
- Pecl GT, Moltschaniwskyj NA (1999) Somatic growth processes: how are they altered in captivity? *Proc Royal Soc London* 266:1133–1139
- Perrin A, Le Bihan E, Koueta N (2004) Experimental study of enriched frozen diet on digestive enzymes and growth of juvenile cuttlefish *Sepia officinalis* L. (Mollusca Cephalopoda). *J Exp Mar Biol Ecol* 311:267–285
- Pinto W, Figueira L, Santos A et al (2013) Is dietary taurine supplementation beneficial for gilthead seabream (*Sparus aurata*) larvae? *Aquaculture* 384–387:1–5
- Pörtner H-O (2010) Oxygen- and capacity-limitation of thermal tolerance: a matrix for integrating climate-related stressor effects in marine ecosystems. *J Exp Biol* 213:881–893
- Quintana D (2009) Valoración de los requerimientos nutricionales de reproductores de pulpo común (*Octopus vulgaris*). PhD Thesis. University of La Laguna, Spain
- Rønnestad I, Thorsen A, Finn RN (1999) Fish larval nutrition: a review of recent advances in the roles of amino acids. *Aquaculture* 177:201–216
- Rønnestad I, Tonheim SK, Fyhn HJ et al (2003) The supply of amino acids during early feeding stages of marine fish larvae: a review of recent findings. *Aquaculture* 227:147–164
- Roura A,G, Redd K et al (2012) Molecular prey identification in wild *Octopus vulgaris* paralarvae. *Mar Biol* 159:1335–1345
- Sargent J, Bell MV, Bell JG et al (1995) Origins and functions of n-3 polyunsaturated fatty acids in marine organisms. In: Phospholipids: Characterization, Metabolism and Novel Biological Applications. Americal Oil Chemical Society, Champaign, p 248–259
- Sargent J, McEvoy LA, Bell JG (1997) Requirements, presentation and sources of polyunsaturated fatty acids in marine fish larval feeds. *Aquaculture* 155:117–127
- Seixas S, Bustamante P, Pierce GJ (2005) Interannual patterns of variation in concentrations of trace elements in arms of *Octopus vulgaris*. *Chemosphere* 59:1113–1124
- Seixas P, Rey-Mendez M, Valente LMP et al (2008) Producing juvenile *Artemia* as prey for *Octopus vulgaris* paralarvae with different microalgal species of controlled biochemical composition. *Aquaculture* 283:83–91
- Seixas P, Rey-Méndez M, Valente LMP et al (2010) High DHA content in *Artemia* is ineffective to improve *Octopus vulgaris* paralarvae rearing. *Aquaculture* 300:156–162
- Semmens JM, Pecl GT, Villanueva R et al (2004) Understanding octopus growth: patterns, variability and physiology. *Mar Freshw Res* 55:367–377
- Sidwell VD, Loomis AL, Foncannon PR, Buzzell DH (1978) Composition of the edible portion of raw (fresh or frozen) crustaceans, finfish, and mollusks. IV. Vitamins. *Mar Fish Rev* 40:1–16
- Sikorski ZE, Kolodziejska I (1986) The composition and properties of squid meat. *Food Chem* 20:213–224
- Sinanoglou VJ, Miniadis-Meimaroglou S (1998) Fatty acid of neutral and polar lipids of (edible) Mediterranean cephalopods. *Food Res Int* 31:467–473
- Sinanoglou VJ, Miniadis-Meimaroglou S (2000) Phospholipids in Mediterranean cephalopods. *Z Naturforsch C* 55:245–255

- Stéphan G, Guillaume J, Lamour F (1995) Lipid-peroxidation in turbot (*Scophthalmus maximus*) tissue: effect of dietary vitamin E and dietary n-6 or n-3 polyunsaturated fatty acids. *Aquaculture* 130:251–268
- Storey KB, Storey JM (1978) Energy metabolism in the mantle muscle of the squid, *Loligo pealeii*. *J Comp Physiol* 123:169–175
- Storey KB, Storey JM (1979) Octopine metabolism in the cuttlefish, *Sepia officinalis*—octopine production by muscle and its role as an aerobic substrate for non-muscular tissues. *J Comp Physiol* 131:311–319
- Storey KB, Storey JM (1983) Carbohydrate metabolism on cephalopod molluscs. In: Hochachka PW (ed) *The Mollusca*. Academic Press Inc., New York, p 91–136
- Storey KB, Storey JM, Johansen K et al (1979) Octopine metabolism in *Sepia officinalis*—effect of hypoxia and metabolite loads on the blood-levels of octopine and related-compounds. *Can J Zool* 57:2331–2336
- Swift K, Johnston D, Moltshaniwskyj N (2005) The digestive gland of the Southern Dumpling Squid (*Euprymna tasmanica*): structure and function. *J Exp Mar Biol Ecol* 315:177–186
- Sykes AV, Domingues PM, Correia M et al (2006) Cuttlefish culture—state of the art and future trends. *Vie Milieu* 56:129–137
- Sykes AV, Almansa E, Lorenzo A et al (2009a) Lipid characterization of both wild and cultured eggs of cuttlefish (*Sepia officinalis* L.) throughout the embryonic development. *Aquac Nut* 15:38–53
- Sykes AV, Oliveira AR, Domingues PM et al (2009b) Assessment of European cuttlefish (*Sepia officinalis*, L.) nutritional value and freshness under ice storage using a developed Quality Index Method (QIM) and biochemical methods. *Food Sci Tech* 42:424–432
- Sykes AV, Pereira D, Rodríguez C et al (2012) Effects of increased tank bottom areas on cuttlefish (*Sepia officinalis*, L.) reproduction performance. *Aquac Res*. doi:10.1111/j.1365–2109.2012.03106.x
- Taylor HH, Anstiss JM (1999) Copper and haemocyanin dynamics in aquatic invertebrates. *Mar Freshw Res* 50:907–931
- Uki N, Kemuyama A, Watanabe T (1986) Optimum protein level in diets for abalone. *Bull Jpn Soc Sci Fish* 52:1005–1012
- Valverde JC, Hernandez MD, Garcia-Garrido S et al (2012) Lipid classes from marine species and meals intended for cephalopod feeding. *Aquac Int* 20:71–89
- Valverde J, Martínez-Llorens S, Vidal A et al (2013) Amino acids composition and protein quality evaluation of marine species and meals for feed formulations in cephalopods. *Aquac Int* 21:413–433
- Van Den Branden CR, Lemaire J et al. (1978) La glande nidamentaire accessoire de *Sepia officinalis* L.: analyses biochimiques des pigments des bactéries symbiotiques. *Annales Soc R Zool Belg* 108:123–139
- Van Den Branden C, Gillis M, Richard A (1980) Carotenoid producing bacteria in the accessory nidamental glands of *Sepia officinalis* L. *Comp Biochem Physiol B* 66:331–334
- Villanueva R, Bustamante P (2006) Composition in essential and non-essential elements of early stages of cephalopods and dietary effects on the elemental profiles of *Octopus vulgaris* paralarvae. *Aquaculture* 261:225–240
- Villanueva R, Riba J, Ruiz-Capillas C et al (2004) Amino acid composition of early stages of cephalopods and effect of amino acid dietary treatments on *Octopus vulgaris* paralarvae. *Aquaculture* 242:455–478
- Villanueva R, Escudero JM, Deulofeu R et al (2009) Vitamin A and E content in early stages of cephalopods and their dietary effects in *Octopus vulgaris* paralarvae. *Aquaculture* 286:277–282
- Vlieg P (1984) Proximate composition of New Zealand squid species. *New Zealand J Sci* 27:45–150
- Vonk HJ (1962) Emulgators in the digestive fluids of invertebrates. *Arch Int Physiol Biochimie* 70:67–85
- Wang T, Hung CCY, Randall DJ (2006) The comparative physiology of food deprivation: from feast to famine. *Annu Rev Physiol* 68:223–251
- Wells MJ, Wells J (1989) Water uptake in a cephalopod and the function of the so-called pancreas. *J Exp Biol* 145:215–226

- Wells MJ, Clarke A (1996) Energetics: the costs of living and reproducing for an individual cephalopod. *Philos Trans Royal Soc London Ser B* 351:1083–1104
- Wolf G, Witters R, Declair W et al (1980) Immunological evidence for hemocyanin-related proteins in mature eggs and embryos of *Sepia officinalis* L. *Arch Int Physiol Bio* 88:B254–B254
- Wolf G, Verheyen E, Vlaeminck A et al (1985) Respiration of *Sepia officinalis* during embryonic and early juvenile life. *Mar Biol* 90:35–39
- Wolfram K, Mark FC, John U et al (2006) Microsatellite DNA variation indicates low levels of genetic differentiation among cuttlefish (*Sepia officinalis* L.) populations in the English Channel and the Bay of Biscay. *Comp Biochem Physiol D* 1:375–383
- Yim M (1978) Développement post-embryonnaire de la glande digestive de *Sepia officinalis* L. (Mollusque Céphalopode). Université de Caen, Caen, p 81
- Zlatanov S, Laskaridis K, Feist C et al (2006) Proximate composition, fatty acid analysis and protein digestibility-corrected amino acid score of three Mediterranean cephalopods. *Mol Nut Food Res* 50:967–970